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Should Zinc be taken with Food?

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Abstract

A recent clinical study reported greater effects of a zinc supplement on indices of essential fatty acid desaturation when the zinc was taken with food, in contrast to a greater plasma zinc response when taken in the fasted state. These results indicate that zinc with food may be more effectively directed toward metabolic functions. This mini review summarizes current knowledge of zinc absorption and metabolism, and the effects of food intake, that may explain these observations.

Keywords:

Supplemental zinc; Plasma zinc; Essential fatty acid metabolism

Introduction

Massih and colleagues recently reported that a zinc supplement, supplying 25 mg of zinc daily as zinc gluconate for two weeks, increased indices of essential fatty acid desaturation when taken with breakfast vs. when taken in the fasted state 30 minutes before breakfast [1]. The dependence of essential fatty acid metabolism on zinc has been previously reported. Early studies associated deficiencies of omega-6 polyunsaturated fatty acids with severe zinc deficiency [2,3]. More recently, a zinc supplement increased indices of erythrocyte membrane fatty acid desaturation and leukocyte fatty acid denaturise 1 mRNA among patients with type 2 diabetes [4]. However, it appears that Massih and colleagues were the first to study the metabolic effects of zinc when taken in the fasted state vs. with food. Given current knowledge on how food intake may moderate zinc metabolism, these findings suggest that zinc with food may be more effectively directed toward zinc-dependent metabolic function, and emphasize the need for further mechanistic study on the dynamics of zinc metabolism.

Zinc Nutrition

Zinc contributes to the structure or function of approximately one tenth of human proteins, in other words, one tenth the molecular machinery behind metabolic and physiological functions [5]. These structural and functional roles provide essential support for numerous metabolic and physiological functions. Essential fatty acid metabolism, vascular and immune functions, and DNA repair, all respond to changes in dietary zinc intake [4,6,9].

In the clinical setting, zinc status is primarily determined by changes in plasma zinc concentration (PZC) [10]. However, due to its tight regulation within the body, PZC varies more within an individual over the course of a day than it does between individuals [11-13]. Except for the diagnosis of severe zinc deficiency, PZC is only recommended as an indicator of population zinc status [10,14].

Part of the need for tight homeostatic control of zinc metabolism is dictated by its lack of a labile storage site in the body. In contrast to micronutrients that may be stored in the body, (e.g., iron, vitamin A, or vitamin D), there are no zinc stores that can be mobilized to maintain zinc nutritional status over periods of low zinc intake [10]. A level of zinc deficiency that leads to impaired functions may thus occur over a relatively short period of low zinc intake.

For example, when healthy men were fed a low zinc diet (4 mg zinc per day, with phytic acid to further inhibit zinc absorption) for two weeks, PZC was conserved. However, increases in double stranded breaks and oxidative damage to DNA were observed, as well as decreases in indices of essential fatty acid desaturation [8,15]. After restoration of dietary zinc by providing 10 mg zinc per day for four weeks, DNA damage and fatty acid metabolic indices were returned to baseline levels. These data demonstrate the potential effects of marginal decreases in dietary zinc on genomic stability and essential fatty acid metabolism over a relatively short period of time. They also highlight the ability of the body to maintain PZC, while zinc-dependent functions decline.

Absorption of dietary zinc

Dietary zinc, solubilized to ionic zinc in the acidic gastric environment, is primarily absorbed in the upper small intestine: the duodenum and proximal jejunum. Zinc absorption is mediated by Zip4, a zinc transport protein localized to the apical membrane of small intestine enterocytes [16,17]. Its absorption is saturable; as the concentration of zinc in the digestate increases, the fraction of zinc absorbed decreases, until a plateau, or transport maximum, is reached [18].

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Zinc, in the form of supplements taken in the post absorptive state after an overnight fast, is rapidly absorbed and increases PZC [19]. In contrast, when zinc is taken with food, the appearance of zinc in plasma is delayed [20]. Some components of food can inhibit zinc absorption. Notably, dietary phytic acid, the storage form of phosphorus in plants, inhibits zinc absorption by forming an insoluble complex with zinc. Calcium joins this complex of phytate and zinc, further reducing zinc absorption [21,22]. The global prevalence of zinc deficiency is associated with the intake of foods high in phytic acid, such as legumes, tubers, whole grains, and seeds [23,24]. However, as protein is added to a meal, a predictable increase in the fractional absorption of dietary zinc is observed, and the capacity of phytic acid to inhibit zinc absorption is reduced [21].

Enterohepatic recirculation

During food intake, a substantial amount of zinc is secreted into the gastrointestinal tract as a component of digestive juices, and it is reabsorbed distally [16,25]. The efficient conservation of this secreted zinc is crucial to survival. Kinetic studies support enterohepatic recirculation of zinc, similar to the recirculation needed for conservation of digestive enzymes [26,27]. The mechanism of enterohepatic zinc recirculation has not been determined in humans. In the ileum, the distal portion of the small intestine where much of the reabsorption of digestive enzymes occurs, the amount of Zip4 ionic zinc transport proteins is not sufficient to account for the reabsorption of digestive zinc. Likewise, passive diffusion, which is observed in animal models at high concentrations of zinc, has not been observed in studies of human zinc absorption [28,29]. Reabsorption of this secreted zinc is therefore most likely mediated by co-transport with another molecule.

Co-absorption of zinc via amino acid transporters could explain the reabsorption of endogenously secreted zinc. Amino acid transporters are localized over the entire length of the small intestine, and cell models support the co-transport of zinc with amino acids [30]. Moreover, several amino acids and related molecules have high affinities for zinc binding [31,32] and their complexes with zinc have the greatest absorption in the ileum compared with other portions of the small intestine [33,34]. The co-absorption of zinc via amino acid transporters may explain the enhancing effect of dietary protein on zinc absorption. Thus, there may be opportunity for zinc from the diet, or a zinc supplement taken with food, to be caught up in enterohepatic recirculation and absorbed distally.

In the absence of food and all the secretions that are stimulated by food (i.e., in the fasted state), the chance of oral zinc getting caught up in this recirculation would be substantially reduced. Essential fatty acid desaturation occurs in most tissues, though primarily in the liver [35,36]. A tendency for supplemental zinc to get caught up in enterohepatic recirculation when taken with food, would naturally direct that zinc to the liver where these cellular metabolic processes take place. Future studies are needed to determine whether zinc cotransport with amino acids, or another mechanism, mediates the enteric recirculation of the zinc contained in digestive secretions.

Effects of food intake on plasma zinc

Interestingly, PZC decreases rapidly after food intake [11,37]. Tracer studies indicate that zinc is directed postprandially into tissues, most likely to the liver [38,39]. However, according to the results of Massih et al., zinc taken in the fasted state, while effective in elevating PZC, does not appear to be directed as efficiently toward essential fatty acid desaturation compared with zinc taken with food [1]. This may be due to the nature of how zinc is utilized: i.e.., as a structural component and cofactor for proteins. The absorption of zinc with amino acids, the building blocks of proteins, may thus be important for the optimal metabolic effects of zinc.

In this case, increases in PZC do not necessarily indicate increased zinc utilization. There is a growing number of examples of demonstrable effects of zinc intake on health outcomes, without detectable changes in PZC [8,40-42]. Further, the lack of correlation between dietary zinc and PZC is a growing theme in zinc clinical research [43]. It is possible that changes in PZC better represent a magnified stress on zinc homeostasis, whether due to severe zinc deficiency, or the absorption of abnormally high amounts of zinc due to the concentration present in the digestive tract. Functional indicators of intracellular zinc, where most zinc-dependent processes occur, therefore, may provide a stronger basis for the development of future clinical indicators of zinc status [10,44,45].

Conclusion

Despite gaps in knowledge of zinc metabolism, a body of scientific evidence is consistent with the concept that zinc may be more effectively directed toward zinc-dependent processes when it is consumed with food. This may be due to coabsorption of some dietary zinc with amino acids, or otherwise entering the pathway for recirculation of endogenously secreted zinc. Furthermore, the kind of food is likely to be important. While foods high in phytic acid inhibit zinc absorption. The functional dependence of zinc on co-absorption with food, how diets may be structured to optimize zinc utilization, and the mechanism of reabsorption of endogenous zinc, represent important areas of future research.

Conflicts of Interest

None

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References

1. Massih YN, Hall AG, Suh J, King JC (2021) Zinc supplements taken with food increase essential fatty acid desaturation indices in adult men compared with zinc taken in the fasted state. J Nutr

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- Horrobin DF, Cunnane SC (1980) Interactions between zinc, essential fatty acids and prostaglandins: relevance to acrodermatitis enteropathica, total parenteral nutrition, the glucagonoma syndrome, diabetes, anorexia nervosa and sickle cell anaemia. Med Hypotheses 6:277-96.
- Koletzko B, Bretschneider A, Bremer HJ (1985) Fatty acid composition of plasma lipids in acrodermatitis enteropathica before and after zinc supplementation. Eur J Pediatr 143:310-4.
- Hernandez MC, Rojas P, Carrasco F, Basfi-Fer K, Valenzuela R et al. (2020) Fatty acid desaturation in red blood cell membranes of patients with type 2 diabetes is improved by zinc supplementation. J Trace Elem Med Biol 62:126571
- 5. Andreini C, Banci L, Bertini I, Rosato A (2006) Counting the zincproteins encoded in the human genome. J Proteome Res 5:196-201.
- Zalewski PD, Beltrame JF, Wawer AA, Abdo AI, Murgia C (2019) Roles for endothelial zinc homeostasis in vascular physiology and coronary artery disease. Crit Rev Food Sci Nutr 59:3511-3525.
- 7. Wessels I, Maywald M,Rink L (2019) Zinc as a gatekeeper of immune function. Nutrients 9: 1286.
- Zyba SJ, Shenvi SV, Killilea DW, Holland TC, Kim E et al. (2017) A moderate increase in dietary zinc reduces DNA strand breaks in leukocytes and alters plasma proteins without changing plasma zinc concentrations. Am J Clin Nutr 105:343-351.
- 9. Mahmoud HM, Ali AF, Al-Timimi DJ (2021) Relationship between zinc status and DNA oxidative damage in patients with type 2 diabetes mellitus. Biol Trace Elem Res 199:1276-1279.
- King JC, Brown KH, Gibson RS, Krebs NF, Lowe NM (2015) Biomarkers of nutrition for development (BOND)-zinc review. J Nutr 146:858S-885S.
- 11. Hambidge KM, Goodall MJ, Stall C, Pritts J (1989) Post-prandial and daily changes in plasma zinc. J Trace Elem Electrolytes Health Dis 3:55-7.
- King JC, Hambidge KM, Westcott JL, Kern DL, Marshall G (1994) Daily variation in plasma zinc concentrations in women fed meals at six-hour intervals. J Nutr 24:508-16.
- 13. Arsenault JE, Wuehler SE, de Romana DL, Penny ME, Sempértegui F et al. (2011) The time of day and the interval since previous meal are associated with plasma zinc concentrations and affect estimated risk of zinc deficiency in young children in Peru and Ecuador. Eur J Clin Nutr 65:184-90.
- 14. Maverakis E, Fung MA, Lynch PJ, Draznin M, Michael DJ et al. (2007) Acrodermatitis enteropathica and an overview of zinc metabolism. J Am Acad Dermatol 56:116-24.
- Suh J, Burke S, Shigenaga M, Killilea D, Holland T et al. (2017) Effect of marginal zinc intake and repletion on essential fatty acid metabolism. FASEB J 31:802.7.
- 16. Krebs NF (2000) Overview of zinc absorption and excretion in the human gastrointestinal tract. J Nutr 130:1374S-7S
- 17. Andrews GK (2008) Regulation and function of Zip4, the acrodermatitis enteropathica gene. Biochem Soc Trans 36:1242-6.
- Hambidge KM, Miller LV, Westcott JE, Sheng X, Krebs NF (2010) Zinc bioavailability and homeostasis Am J Clin Nutr 91:1478S-1483S.
- 19. Wessells KR, Jorgensen JM, Hess SY, LR Woodhouse, Peerson JM et al. (2010) Plasma zinc concentration responds rapidly to the initiation and discontinuation of short-term zinc supplementation in healthy men J Nutr 40:2128-33.

- Nève J, Hanocq M, Peretz A, Khalil FA, Pelen F (1992) Absorption and metabolism of oral zinc gluconate in humans in fasting state, during, and after a meal. Biol Trace Elem Res 32:201-12.
- 21. Miller LV, Krebs NF, Hambidge KM (2013) Mathematical model of zinc absorption: effects of dietary calcium, protein and iron on zinc absorption. Br J Nutr Feb 28; 109:695-700.
- 22. Kratzer FH, Vohra P (1986) Chelates in Nutrition. Boca Raton pages:1-178.
- Dahdouh S, Grande F, Espinosa SN, Vincent A, Gibson R et al. (2019) Development of the FAO/INFOODS/IZINCG global food composition database for phytate. J Food Compost Anal 78:42-48.
- 24. Wessells KR, Brown KH (2012) Estimating the global prevalence of zinc deficiency: results based on zinc availability in national food supplies and the prevalence of stunting. PLoS One 7:e50568.
- Matseshe JW, Phillips SF, Malagelada JR, McCall JT (1998) Recovery of dietary iron and zinc from the proximal intestine of healthy man: studies of different meals and supplements. Am J Clin Nutr 33:1946-53.
- J Nève , M Hanocq, A Peretz, F Abi Khalil, F Pelen, et al. (1991) Pharmacokinetic study of orally administered zinc in humans: evidence for an enteral recirculation. Eur J Drug Metab Pharmacokinet 16:315-23.
- 27. Rothman S, Liebow C, Isenman L (2002) Conservation of digestive enzymes. Physiol Rev 82:1-18.
- Menard MP, Cousins RJ (1983) Effect of citrate, glutathione and picolinate on zinc transport by brush border membrane vesicles from rat intestine. J Nutr 113:1653-6.
- Miller LV, Krebs NF, Hambidge KM (2007) A mathematical model of zinc absorption in humans as a function of dietary zinc and phytate. J Nutr 137:135-4.
- Sauer AK, Pander S, Hagmeyer S, Tarana L, Mattes AK et al. (2017) Characterization of zinc amino acid complexes for zinc delivery in vitro using Caco-2 cells and enterocytes from hiPSC. Biometals 30:643-661.
- 31. Krężel A, Maret W (2016) The biological inorganic chemistry of zinc ions. Arch Biochem Biophys 611:3-19.
- Trzaskowski B, Adamowicz L, Deymier PA (2008) A theoretical study of zinc(II) interactions with amino acid models and peptide fragments. J Biol Inorg Chem 13:133-7.
- Johnson WT, Evans GW (1982) Tissue uptake of zinc in rats following the administration of zinc dipicolinate or zinc histidinate. J Nutr 112:914-9.
- Wapnir RA, Khani DE, Bayne MA, Lifshitz F (1983) Absorption of zinc by the rat ileum: effects of histidine and other low-molecularweight ligands. J Nutr 113:1346-54.
- Leonard AE, Kelder B, Bobik EG, LT C, Parker-Barnes JM (2000) cDNA cloning and characterization of human Delta5-desaturase involved in the biosynthesis of arachidonic acid. Biochem J Pt 3:719-24.
- Nakamura MT, Nara TY (2003) Essential fatty acid synthesis and its regulation in mammals. Prostaglandins Leukot Essent Fatty Acids 68:145-50.
- Wallock LM, King JC, Hambidge KM, English-Westcott JE, Pritts J, et al.(1993) Meal-induced changes in plasma, erythrocyte, and urinary zinc concentrations in adult women. Am J Clin Nutr 58:695-701.

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- Lowe NM, Woodhouse LR, King JC (1998) A comparison of the short-term kinetics of zinc metabolism in women during fasting and following a breakfast meal. Br J Nutr 80:363-70.
- DeGrado TR, Kemp BJ, Pandey MK, Jiang H, Gunderson TM et al. (2016) First PET Imaging Studies With 63Zn-Zinc Citrate in Healthy Human Participants and Patients With Alzheimer Disease. Mol Imaging 9;15.
- Consolo LZZ, Melnikov P, Cônsolo FZ, Nascimento VA, Pontes JV (2013)Zinc supplementation in children and adolescents with acute leukemia. Eur J Clin Nutr 67:1056-9.
- 41. Ariff S, Krebs NF, Soofi S, Westcott J, Bhatti Z (2014) Absorbed zinc and exchangeable zinc pool size are greater in Pakistani infants receiving traditional complementary foods with zinc-fortified micronutrient powder. J Nutr 144:20-6.
- Radhakrishna KV, Hemalatha R, Babu Geddam JJ, Ajey Kumar P, Balakrishna N (2013) Effectiveness of Zinc Supplementation to Full Term Normal Infants: A Community Based Double Blind, Randomized, Controlled, Clinical Trial. PLoS One 8: e61486.
- 43. King JC (2018) Yet Again, Serum Zinc Concentrations Are Unrelated to Zinc Intakes. J Nutr 148: 1399–1401.
- Van Wouwe JP (1995) Clinical and laboratory assessment of zinc deficiency in Dutch children. A review. Biol Trace Elem Res 49:211-25.
- 45. Van Wouwe JP, Veldhuizen M, De Goeij JJ, Van den Hamer CJ (1991) Laboratory assessment of early dietary, subclinical zinc deficiency: a model study on weaning rats. Pediatr Res 29:391-5.